

Relationships between leaf chlorophyll content and spectral reflectance and algorithms for non-destructive chlorophyll assessment in higher plant leaves

Anatoly A. Gitelson^{1,2} *, Yuri Gritz † ², Mark N. Merzlyak³

Received July 10, 2002 · Accepted August 17, 2002

Summary

Leaf chlorophyll content provides valuable information about physiological status of plants. Reflectance measurement makes it possible to quickly and non-destructively assess, in situ, the chlorophyll content in leaves. Our objective was to investigate the spectral behavior of the relationship between reflectance and chlorophyll content and to develop a technique for non-destructive chlorophyll estimation in leaves with a wide range of pigment content and composition using reflectance in a few broad spectral bands. Spectral reflectance of maple, chestnut, wild vine and beech leaves in a wide range of pigment content and composition was investigated. It was shown that reciprocal reflectance $(R_{\lambda})^{-1}$ in the spectral range λ from 520 to 550 nm and 695 to 705 nm related closely to the total chlorophyll content in leaves of all species. Subtraction of near infra-red reciprocal reflectance, $(R_{NIR})^{-1}$, from $(R_{\lambda})^{-1}$ made index $[(R_{\lambda})^{-1} - (R_{NIR})^{-1}]$ linearly proportional to the total chlorophyll content in spectral ranges λ from 525 to 555 nm and from 695 to 725 nm with coefficient of determination $r^2 > 0.94$. To adjust for differences in leaf structure, the product of the latter index and NIR reflectance $[(R_{\lambda})^{-1} - (R_{NIR})^{-1}]^*(R_{NIR})$ was used; this further increased the accuracy of the chlorophyll estimation in the range λ from 520 to 585 nm and from 695 to 740 nm. Two independent data sets were used to validate the developed algorithms. The root mean square error of the chlorophyll prediction did not exceed 50 µmol/m² in leaves with total chlorophyll ranged from 1 to $830 \,\mu\text{mol/m}^2$.

Key words: chlorophylls - non-destructive assessment - leaf optics - reflectance

Abbreviations: Chl = Chlorophyll. - NIR = Near Infra Red. - RMSE = Root Mean Square Error

¹ Center for Advanced Land Management Information Technologies, School of Natural Resource Sciences, University of Nebraska-Lincoln, 113 Nebraska Hall, Lincoln, NE 68588 – 0517, USA

² J. Blaustein Institute for Desert Research, Ben-Gurion University of the Negev, Sede-Boker Campus 84993, Israel

³ Department of Physiology of Microorganisms, Faculty of Biology, Moscow State University, 119899 GSP, Moscow W-234, Russia

^{*} E-mail corresponding author: gitelson@calmit.unl.edu

Introduction

The chlorophylls, Chl *a* and Chl *b*, are virtually essential pigments for the conversion of light energy to stored chemical energy. The amount of solar radiation absorbed by a leaf is a function of the photosynthetic pigment content; thus, chlorophyll content can directly determine photosynthetic potential and primary production (Curran et al. 1990, Filella et al. 1995). In addition, Chl gives an indirect estimation of the nutrient status because much of leaf nitrogen is incorporated in chlorophyll (Filella et al. 1995, Moran et al. 2000). Furthermore, leaf chlorophyll content is closely related to plant stress and senescence (Hendry 1987, Merzlyak and Gitelson 1995, Peñuelas and Filella 1998, Merzlyak et al. 1999).

Traditionally, leaf extraction with organic solvents and spectrophotometric determination in solution is required for pigment analysis with wet chemical methods (e.g., Lichtenthaler 1987). Recently, alternative solutions of leaf pigment analysis (i.e., chlorophyll, carotenoids and anthocyanins) with non-destructive optical methods have been developed. These newer methods are non-destructive, inexpensive, quick and now possible in the field (Buschmann and Nagel 1993, Gitelson and Merzlyak 1994 a, b, Markwell et al. 1995, Gamon and Surfus 1999, Gitelson et al. 2001, 2002).

Transmittance and reflectance spectroscopy is applied extensively for non-destructive estimation of leaf ChI (Adams and Arkin 1977, Aoki et al. 1986, Curran et al. 1990, Gitelson and Merzlyak 1994 a, b, 1996, 1977, Markwell et al. 1995, Gitelson et al. 1996, Blackburn 1998, Datt 1998, Gamon and Surfus 1999, Richardson et al. 2002). Relationships between reflectance in the visible range and leaf Chl content are essentially nonlinear (e.g., Buschmann and Nagel 1993, Gitelson and Merzlyak 1994 a). First-difference transformation of the apparent absorbance, the logarithm of reciprocal reflectance (R)⁻¹, was found to be the best predictor for nitrogen and chlorophyll in fresh bigleaf maple leaves (Yoder and Pettigrew-Crosby 1995). Gitelson and Merzlyak (1994 a and b) have found that reflectances in the spectral bands located quite far from the main absorption bands of pigments near 550 nm and 700 nm were closely hyperbolically related to Chl for a variety of plant species and in a wide range of pigment content and composition. Indices based at these spectral bands were proposed and used to estimate ChI content in the leaves of various plant species (Chappele et al. 1992, Gitelson and Merzlyak 1994 a, b, 1996, 1997, Gitelson et al. 1996, Lichtenthaler et al. 1996, Gamon and Surfus 1999).

Richardson et al. (2002) evaluated both non-destructive absorbance and reflectance methods for Chl assessment. They compared the performance of two commercially available hand-held Chl absorbance meters with that of several reflectance indices for the estimation of leaf-level Chl and found that some indices based on reflectance in the red edge region (Gitelson and Merzlyak 1994 a) were much better indicators of Chl content than some of the more commonly used indices. The best reflectance indices were found to be better

indicators of Chl compared with those based on absorbance measurements (Richardson et al. 2002).

Sims and Gamon (2002) examined hundreds of leaves of non-related plant species and proved that reflectance in the spectral channel around 700 nm was the most sensitive indicator of ChI and that indices R_{750}/R_{700} and $(R_{750}-R_{705})/(R_{750}+R_{705}),$ Gitelson and Merzlyak (1994 a), could be used as a measure of ChI content. However, the correlation of these indices with ChI was weaker when applied across a wide range of species. The modified indices $(R_{750}-R_{445})/(R_{700}-R_{445})$ and $(R_{NIR}-R_{705})/(R_{NIR}+R_{705}-2R_{445}),$ which were developed to eliminate the effect of variability in surface reflectance between species by incorporating reflectance at 445 nm, R_{445} , produced substantially better correlation with total ChI content.

Although a leaf consisting of cuticula, epidermis, palisade and spongy parenchyma, etc. with numerous boundaries and containing high amounts of pigments is very complicated from an optical point of view (Gates et al. 1965, Heath 1969, Fukshansky 1981, Vogelmann 1993, Richter and Fukshansky 1996), it was found that reciprocal reflectance alone at certain wavelengths could be used for chlorophyll quantification (Gitelson et al. 1996). Based on that finding, algorithms for nondestructive estimation of carotenoids (Gitelson et al. 2002) and anthocyanins (Gitelson et al. 2001) were developed in which reciprocal reflectances at 550 nm and 700 nm were used to eliminate the Chl contribution in spectral ranges with overlapping absorption by Chl and other pigments. However, the mechanisms responsible for close relationships between reciprocal reflectance and pigment content needs more detailed investigations.

The indices for chlorophyll estimation based on reflectances in narrow spectral bands (usually 1 to 2 nm wide) at 550 nm and/or around 700 nm have been tested (Aoki et al. 1986, Gitelson and Merzlyak 1997, Datt 1998, Gamon and Surfus 1999, Carter and Knapp 2001, Richardson et al. 2002, Sims and Gamon 2002) and showed good performance in Chl estimation. However, to provide sufficient sensitivity to a small variation of Chl, broad spectral range is required. It is also desirable to use an algorithm that is minimally sensitive to the differences in leaf structure to avoid species-specific calibration if the estimation of absolute Chl content is desired. We are not aware of any studies that have attempted to assess how the accuracy of Chl estimation depends on the width of spectral bands and how wide spectral bands must be to provide the required accuracy.

The goal of this research was to investigate the spectral behavior of the relationship between reflectance and ChI content and to develop a technique for ChI estimation in leaves with a wide range of pigment content and composition using reflectance in a few broad spectral bands. The first step was to find spectral ranges where ChI related closely to reciprocal reflectance (R_{λ})⁻¹ for all species studied. We found that reciprocal reflectances in narrow bands in the green and red edge spectral regions were sensitive and closely related to the ChI

content. Subtraction of $(R_{NIR})^{-1}$ from $(R_{\lambda})^{-1}$ made index $(R_{\lambda})^{-1}$ ($R_{NIR})^{-1}$ linearly proportional to the total ChI content in wide spectral bands 525 to 555 nm and 695 to 725 nm. To correct for differences in leaf structure, we suggested the product of the above index and NIR reflectance $[(R_{\lambda})^{-1}\text{-}(R_{NIR})^{-1}] \star R_{NIR}$. This algorithm made it possible to reach a high accuracy of ChI estimation with Root Mean Square Error (RMSE) < 40 $\mu\text{mol/m}^2$ in broad spectral bands (520–585 nm and 695–735 nm). Finally, algorithms were validated by independent data sets.

Materials and Methods

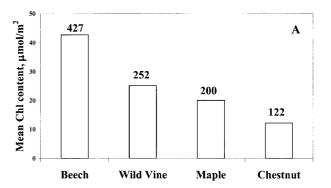
Juvenile, mature and senescent leaves of Norway maple (*Acer platanoides* L.) and horse chestnut (*Aesculus hippocastanum* L.) were collected in a park at Moscow State University in the spring, summer and fall of 1992–2000. Second-flush beech leaves (*Fagus sylvatica* L.) grown on the University of Karlsruhe campus were taken in August and September 1996 and in August 2000, and wild vine shrub (*Parthenocissus tricuspidata* L.) leaves were collected in August and September 1996. Leaves were visually selected according to their difference in color. Leaves healthy and homogeneous in color without anthocyanin pigmentation or visible symptoms of damage were used in the experiments.

The leaf pigment content was determined from the same leaf samples used for reflectance measurement. Circular pieces were cut from the leaves and extracted with 100% acetone or methanol using a mortar. The pigment extracts were centrifuged for 3–5 min in glass tubes to make the extract fully transparent. The resulting extracts were immediately assayed spectrophotometrically. Specific absorption coefficients of Chl *a*, Chl *b* and total carotenoids reported by Lichtenthaler (1987) were used. The accepted molecular weight of carotenoids was 570.

Adaxial reflectance (R) and transmittance (T) spectra of the leaves were taken in a spectral range between 400 and 800 nm with a spectral resolution of 2 nm with a Hitachi 150-20 spectrophotometer

Table 1. Pigment content and composition in leaves used for model development and model validation. Pigment content in μ mol/m², n is the number of leaves in each data set.

Data Set	n	Total Chlorophyll	Carotenoids
Model development			
Beech - 1996	38	97-832	53-240
Wild Vine	19	47-530	21-180
Maple - 92-98	65	1-570	29-170
Chestnut	20	10-470	53-170
	142	1-832	21-240
Model validation			
Maple - 1999	23	1-460	16-120
Beech – 2000	34	14-670	36-210
	57	1-670	16-210
Total	199	1-832	16-240



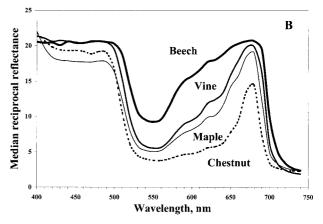


Figure 1. (A) Mean Chl content in four leaf groups used for model development. (B) Median reciprocal reflectance spectra for each leaf group. Maximal reciprocal reflectance in the range from 500 nm to 700 nm was in beech leaves with maximal Chl content, and minimal reciprocal reflectance was in chestnut leaves with minimal Chl. In the blue range, reciprocal reflectance depends on both chlorophylls and carotenoids content.

(measurements of maple and chestnut leaves) and a Shimadzu 2101 PC spectrophotometer (measurements of beech and wild vine leaves) equipped with an integrating sphere. Leaf reflectance spectra were recorded against barium sulphate as a standard; black velvet was used as a background.

Four data sets containing 140 leaves (maple 1992–1998, chestnut, beech 1996 and wild vine) were used for model development. Two independent data sets (23 maple-1999 and 34 beech-2000 leaves) were used to validate the models. Relationships between the developed indices and ChI content obtained for the model development data sets were inverted, and reflectance data from the model validation data sets were used to calculate the predicted ChI content. Finally, the predicted ChI content was compared with analytically measured ChI values, and the RMSE of the ChI prediction was determined.

Results

Pigment content and composition

Pigment content and composition in the leaves varied widely (Table 1). Carotenoids were the dominant pigments in leaves

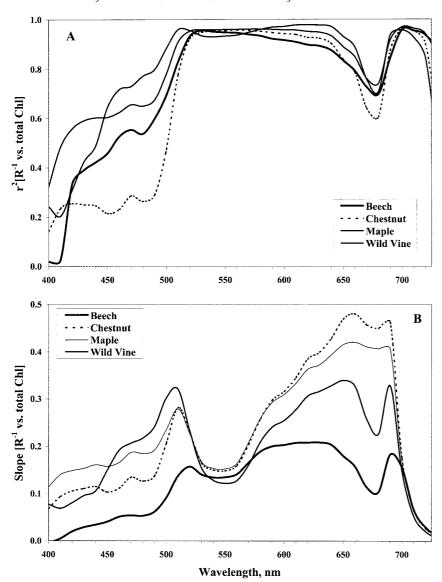


Figure 2. (A) Spectra of the coefficient of determination, r^2 , and (B) the slope of the linear relationship between reciprocal reflectance $(R_{\lambda})^{-1}$ and total ChI content for four leaf group studied (Table 1: model development data sets).

with a total pigment content (chlorophylls + carotenoids) of less than $50\,\mu\text{mol/m}^2$; they constituted more than $97\,\%$ of the total pigment content. With an increase in total pigment content, the carotenoids fraction decreased sharply; Chl presented the major portion of pigments in the leaves with a total pigment content > $200\,\mu\text{mol/m}^2$. With a further increase in leaf greenness, the ratio of carotenoids to total pigment content decreased only a little: from $30\,\%$ in slightly green leaves to $20\,\%$ in dark-green leaves. The relationship between carotenoids and total Chl was linear with a coefficient of determination $r^2 = 0.87$.

Leaves of four plant species, used for model development, were quite different with respect to total ChI content. Mean ChI content ranged from a little bit higher than $120\,\mu\text{mol/m}^2$ in chestnut leaves to more than $400\,\mu\text{mol/m}^2$ in beech leaves (Fig. 1A).

Spectral characteristics of leaves and their relations with total ChI content

The difference in mean Chl content in the four leaf groups studied manifested itself in different reciprocal reflectance, $(R)^{-1}$, spectra (Fig. 1B). In the blue range from 400 to 500 nm, $(R)^{-1}$ was the highest in beech and wild vine leaves and slightly lower in chestnut and maple. In this range, $(R)^{-1}$ was affected by both Chl and carotenoids. Considerable changes of $(R)^{-1}$ occurred in the range between 500 and 700 nm. With an increase in Chl content, both green edge (around 530 nm) and red edge (around 700 nm) shifted toward longer wavelengths and the reciprocal reflectance increased. The highest reciprocal reflectance was in beech leaves with maximal mean Chl, and the lowest one was in chestnut leaves with minimal mean Chl.

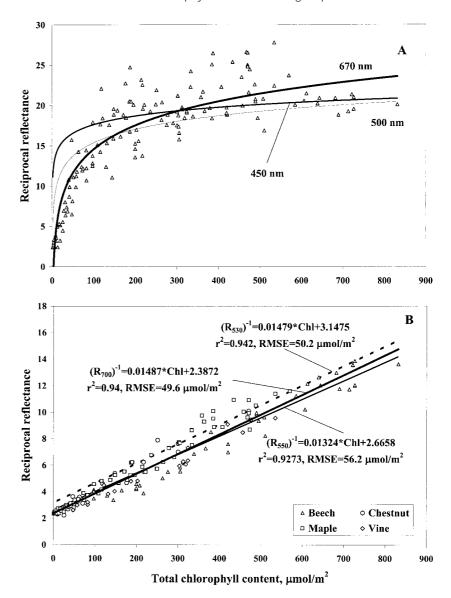


Figure 3. Relationship between reciprocal reflectance and total chlorophyll content (A) in the blue (450 and 500 nm) and the red (670 nm), and (B) in the green (530 and 550 nm) and red edge (700 nm).

To find spectral bands where reciprocal reflectance was sensitive to total ChI content, we studied the spectral behavior of the coefficient of determination, r^2 , and the slope of the linear relationship $(R_{\lambda})^{-1}$ vs. total ChI content (Fig. 2). In the blue (400–500 nm) and red (around 680 nm), r^2 was minimal (Fig. 2 A). In these spectral ranges, the relationship $(R_{\lambda})^{-1}$ vs. total ChI was essentially non-linear (Fig. 3 A). For ChI < $100\,\mu\text{mol/m}^2$, the sensitivity of $(R_{\lambda})^{-1}$ to ChI was high; for ChI > $150\,\mu\text{mol/m}^2$, the relationship leveled off, and $(R_{\lambda})^{-1}$ became virtually insensitive to ChI. In the spectral range between 510 and 620 nm and near 700 nm, the function $(R_{\lambda})^{-1}$ vs. ChI was linear for all species with r^2 > 0.9 (Figs. 2 A and 3 B).

The slope of the relationship $(R_{\lambda})^{-1}$ vs. Chl reached maximal values around 510-520 nm, at 650 nm and at 690-695 nm, spectral ranges located far from main peaks of Chl absorption (Fig. 2 B). In the blue range, the slope depended

upon the content of ChI and carotenoids and varied widely between species with a minimal slope for beech, which had a maximal mean ChI content. In the orange and red ranges, the slopes were inversely related to mean ChI content: the higher ChI content, the lower the slope; so, the absorption per ChI unit was higher for species with lower ChI than for species with higher ChI. In the green (between 520 and 580 nm) and red edge (around 700 nm), the slopes of the relationships $(R_3)^{-1}$ vs. ChI remained fairly similar for different species.

The high linear correlation between Chl content and $(R_{\lambda})^{-1}$ in a wide range from 510 to 650 nm and in a narrow range around 700 nm (Figs. 2 A and 3 B) suggested the use of $(R_{\lambda})^{-1}$ as a measure of total Chl content. However, to be applied for variety of species, relationship $(R_{\lambda})^{-1}$ vs. Chl should have minimal variation in the slope between species. The coefficient of variation of the slope between species was the highest (more

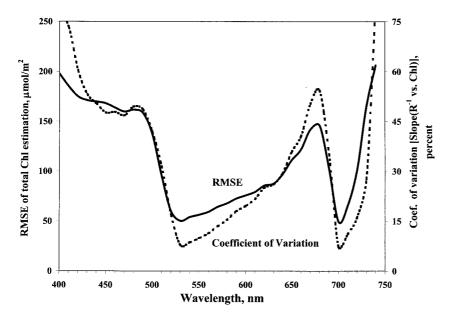


Figure 4. The coefficient of variation of the slope of the linear relationship between $(R_{\lambda})^{-1}$ and total ChI content (right axis), and the RMSE of ChI estimation by reciprocal reflectance (left axis) for the four species studied.

than 45%) in the blue range between 400 and 490 nm and in the red range near 675 nm (Fig. 4). It reached minimal values (below 10%) in the green range between 520 and 580 nm, and in the red edge around 700 nm; in these spectral ranges, the RMSE of ChI estimation had minimal values (50 to $60\,\mu\text{mol}$).

Thus, the relationship $(R_{\lambda})^{-1}$ vs. ChI showed several notable spectral features.

- 1. In the main bands of pigment absorption, the blue and the red, the relationships were essentially non-linear. $(R_{\lambda})^{-1}$ was sensitive to Chl < 150 μ mol/m², but insensitive to moderate-to-high Chl.
- 2. In the blue and the red, the slopes of the relationship (R_λ)⁻¹ vs. ChI varied widely between species. In the green and red edge, the slopes remained nearly the same for all species studied. In these spectral ranges, reciprocal reflectance was a measure of ChI content, and the RMSE of the ChI estimation had its minimal values.

Algorithm development and validation

For leaves containing trace amounts of ChI, reciprocal reflectance remained significantly above zero (Fig. 3 B). In the range beyond 550 nm, the intercept (R_{λ})⁻¹ vs. ChI was almost equal to (R_{NIR})⁻¹ (not shown). Thus, we subtracted the reciprocal reflectance in the NIR range, (R_{NIR})⁻¹, from (R_{λ})⁻¹ to make the index

$$[(R_{\lambda})^{-1}-(R_{NIR})^{-1}]$$

linearly proportional to total Chl. At 700 nm, the intercept of the relationship $[(R_{700})^{-1}-(R_{NIR})^{-1}]$ vs. Chl dropped 16-fold and became very close to zero. The intercept $[(R_{550})^{-1}-(R_{NIR})^{-1}]$ vs. Chl also decreased significantly (six-fold), but nevertheless, remained positive (Fig. 5 A); it became negligible beyond

 $550\,\mathrm{nm}$. Remarkably, the subtraction of $(\mathrm{R_{NIR}})^{-1}$ made it possible to extend significantly the spectral range of an accurate ChI estimation. It was especially evident in the wide ranges from 525 to $555\,\mathrm{nm}$ and 695 to $735\,\mathrm{nm}$. The RMSE of ChI estimation by indices

$$[(R_{525-555})^{-1} - (R_{NIR})^{-1}]$$
$$[(R_{695-725})^{-1} - (R_{NIR})^{-1}]$$

was below $50 \,\mu\text{mol/m}^2$ (Fig. 5 B).

A further increase in the accuracy of ChI estimation was achieved by using the product of the index $[(R_{\lambda})^{-1}-(R_{NIR})^{-1}]$ and R_{NIR} :

$$[(R_{\lambda})^{-1} - (R_{NIR})^{-1}] * R_{NIR} = (R_{NIR}/R_{\lambda}) - 1.$$

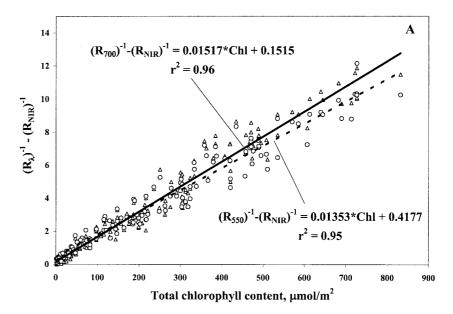
This algorithm is slightly different from R_{NIR}/R_{550} and R_{NIR}/R_{700} , suggested by Gitelson and Merzlyak (1994 a, b). Importantly, however, the intercept of the function $(R_{NIR}/R_{\lambda})^{-1}$ vs. ChI was quite close to zero, which made the index linearly proportional to the ChI content. The main differences in the spectral behavior of all three indices, $(R_{\lambda})^{-1}$, $[(R_{\lambda})^{-1} - (R_{NIR})^{-1}]$ and (R_{NIR}/R_{λ}) -1, were in the green and red edge ranges (Fig. 6). The RMSE of ChI estimation by the indices

$$(R_{NIR}/R_{520-585})-1$$

 $(R_{NIR}/R_{695-740})-1$

was minimal among indices, remaining below 50 μmol/m².

Validation of the proposed technique has been carried out for two independent data sets: maple-1999 and beech-2000 with a wide variation of pigment content (Table 1). The relationship index vs. Chl obtained for model development data sets was inverted, and reflectance data from the model validation data sets were used to calculate the predicted Chl content. The predicted Chl content was compared with ana-



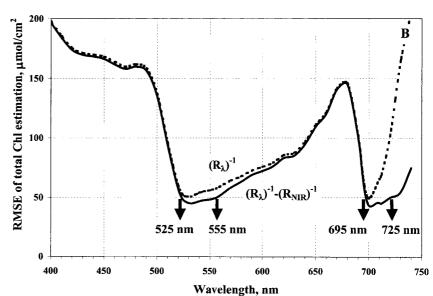


Figure 5. (A) Relationship between index $[(R_{\lambda})^{-1} - (R_{NIR})^{-1}]$ and total ChI content in the green (550 nm) and red edge (700 nm) regions for the four leaf groups studied; (B) RMSE of total ChI estimation by $(R_{\lambda})^{-1}$ and index $[(R_{\lambda})^{-1} - (R_{NIR})^{-1}]$. Spectral ranges where the RMSE of total ChI estimation dropped below $50 \, \mu \text{mol/m}^2$ are shown.

lytically measured ChI, and the RMSE of the ChI prediction was determined (Table 2). For all 57 leaves examined, the RMSE of the total ChI prediction by index (R_{NIR}/R_{λ})-1 in spectral ranges λ from 520 to 585 and from 695 to 740 nm was less than $49\,\mu\text{mol/m}^2$.

Discussion

In the blue and red spectral ranges, specific absorption coefficients of pigments are very high (e.g., Heath 1969, Lichtenthaler 1987) and the depth of light penetration into the leaf is very low (Kumar and Silva 1973, Cui et al. 1991, Vogelmann et al. 1991, Vogelmann 1993, Fukshansky et al. 1993, Merzlyak and Gitelson 1995). As a result, even low amounts of pig-

ments are sufficient to saturate absorption. When Chl exceeded 150 μ mol/m², total absorption reached maximal values (more than 90 %), the depth of light penetration drastically decreased, and a further increase in pigment content did not cause an increase in total absorption. Thus, the relationship $(R_{\lambda})^{-1}$ vs. total Chl leveled off, and $(R_{\lambda})^{-1}$ became virtually insensitive to Chl (see also, Thomas and Gausman 1977, Gausman 1982, Chapelle et al. 1992, Buschmann and Nagel 1993, Gitelson and Merzlyak 1994 a, b, 1997, Gamon and Surfus 1999). In the spectral regions located closer to the main absorption bands, the slope $(R_{\lambda})^{-1}$ vs. Chl depended strongly on mean pigment content in leaf group (Fig. 2B). The closer the spectral band was to the main absorption band of pigments, the lower the Chl content at which saturation of $(R_{\lambda})^{-1}$ vs. Chl relationship appeared.

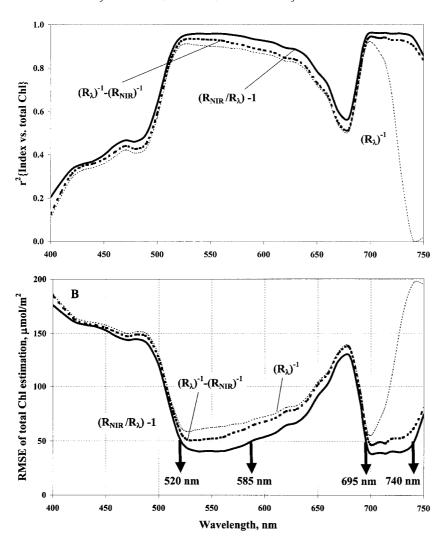


Figure 6. (A) The coefficient of determination r^2 and (B) RMSE of total ChI estimation by three indices proposed for ChI assessment: $(R_{\lambda})^{-1}$, $[(R_{\lambda})^{-1}, (R_{NIR})^{-1}]$, and (R_{NIR}/R_{λ}) -1. Spectral ranges where RMSE of ChI assessment dropped below $50 \, \mu \text{mol}/m^2$ are shown.

Table 2. Root mean square error of leaf chlorophyll prediction (in μmol/ m^2) for model validation data sets: beech–2000 and maple–1999. The relationship index vs. Chl obtained for model development data sets was inversed, and reflectance data from the model validation data sets were used to calculate the predicted Chl content. Then, the predicted Chl content was compared with analytically measured actual Chl values, and the RMSE of Chl prediction was determined. Note: Chl content in beech leaves ranged from 14 to 670 μmol/ m^2 ; whereas, it ranged from 1 to 460 μmol/ m^2 in maple leaves.

Waveband λ (nm)	Index	Beech-2000	Maple - 1999	
520 – 550	$(R_{\lambda})^{-1}$	63	59	
525 – 555	$(R_{\lambda})^{-1}$ - $(R_{NIR})^{-1}$	59	49	
520 – 585	(R_{NIR}/R_{λ}) -1	48	35	
695 – 705	$(R_{\lambda})^{-1}$	64	58	
695 – 725	$(R_{\lambda})^{-1}$ - $(R_{NIR})^{-1}$	61	49	
695 – 740	$(R_{NIR}/R_{\lambda}) - 1$	49	33	

In the green and in the red edge (around 700 nm), specific absorption coefficient of chlorophylls in extract is very low; it does not exceed even 6 % of that in the blue and red (e.g., Heath 1969, Lichtenthaler 1987); however, green leaves absorb more than 80 % of incident light in these spectral ranges (e.g., Moss and Loomis 1952, Heath 1969, Gausman et al. 1969, Gausman and Allen 1973, Gitelson and Merzlyak 1994 a). In these spectral ranges, depth of light penetration into the leaf was found to be four- to six-fold higher than in the blue and red (e.g., Fukshansky et al. 1993, Fig. 2 in Merzlyak and Gitelson 1995). Therefore, in the green and red edge absorption of light is high enough to provide high sensitivity of $(R_{\lambda})^{-1}$ to Chl content and much lower than in the blue and red to avoid saturation of the $(R_{\lambda})^{-1}$ vs. Chl relationship for moderate to high Chl.

The Kubelka-Munk remission function, which is proportional to the ratio of the absorption coefficient, k, to the scat-

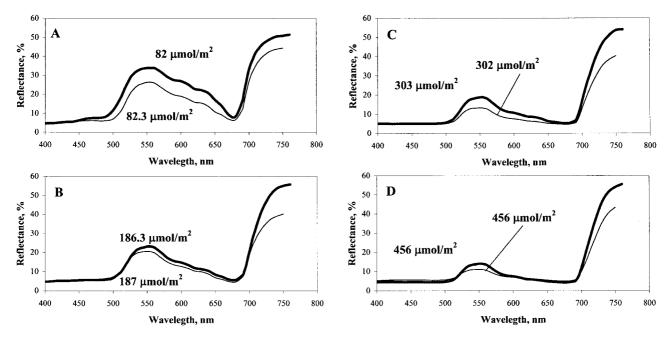


Figure 7. Reflectance spectra of leaves with almost the same total ChI content (indicated as numbers near the curves) and different NIR reflectance. For the same ChI content, higher NIR reflectance values correspond to higher reflectance in the green (around 550 nm) and in the range of the red edge (near 700 nm).

tering coefficient, s (e.g. Wendlandt and Hecht 1966) relates very closely to leaf reciprocal reflectance (Appendix 1). Therefore, in the green and red edge ranges, the ratios (k/s)_{green} and (k/s)_{red edge} were closely and linearly related to the total Chl content (Fig. 3 B). The k/s ratio per total Chl units (specific ratio in m^2/μ mol) remained virtually invariant (0.0129 \pm 0.003 m^2/μ mol) in leaves with Chl from 1 to 830 μ mol/ m^2 and a wide range of pigment composition.

In an attempt to understand why the indices (R_{NIR}/R₅₅₀)-1 and (R_{NIR}/R₇₀₀)-1 were effective in improving the accuracy of Chl assessment, we compared reflectance spectra of leaves with almost the same ChI content but different R_{NIR}. We found with no exception that the higher the reflectance in the NIR range, the higher the reflectances in the range around 550 nm and 700 nm (Fig. 7). The processes that govern behavior of reflectance in the NIR range and the visible spectrum are very different. In the NIR range, an increase in reflectance might be caused by an increase in leaf thickness or/and density; in the visible range, an increase in reflectance indicates a decrease in pigment content (surface reflectance of adaxial surface for all leaves studied was virtually the same). In leaves with the same Chl content, an increase in leaf thickness might lead to an increase in $R_{\mbox{\scriptsize NIR}}$ and to a decrease in Chl concentration (per volume); the latter caused an increase in reflectances in the green and red edge that was observed (Fig. 7). A decrease in leaf thickness might cause a decrease in R_{NIR} and an increase of ChI concentration, which caused a decrease in reflectance in the green and red edge. With a variation in leaf thickness, reflectances $(R_{520-585})^{-1}$, $(R_{695-740})^{-1}$, and R_{NIR} varied in the same direction, and, thus, indices $(R_{NIR}/R_{520-570})$ -1 and $(R_{NIR}/R_{695-735})$ -1 became less sensitive to this factor. As a result, these indices correlated with total ChI much more closely than $(R_{520-570})^{-1}$ and $(R_{700})^{-1}$ (Fig. 6).

Leaf surface reflectance virtually did not change in leaves studied, so, indices worked accurately across the species. In the case when surface reflectance varies, incorporating reflectance in the blue range makes it possible to eliminate the effect of surface reflectance (Sims and Gamon 2002). We recommend using the reflectance in the range between 430 and 470 nm in modified indices:

$$\begin{split} &[(R_{520-555}-R_{440-480})^{-1}\text{-}(R_{NIR}-R_{440-480})^{-1}]\\ &[(R_{695-725}-R_{440-480})^{-1}\text{-}(R_{NIR}-R_{440-480})^{-1}]\\ \text{and}\\ &[(R_{NIR}-R_{440-480})/(R_{520-580}-R_{440-480})]\text{-}1\\ &[(R_{NIR}-R_{440-480})/(R_{695-740}-R_{440-480})]\text{-}1 \end{split}$$

Comparison with previously developed indices

Using our data sets, we tested the accuracy of the indices that had been previously developed for ChI estimation and compared it with the performance of the indices proposed in this paper. In Figure 8, relationships between total chlorophyll content and the following indices are presented:

$$(R_{800}-R_{680})/(R_{800}+R_{680})$$
 – Blackburn 1998, (R_{800}/R_{680}) – Blackburn 1998,

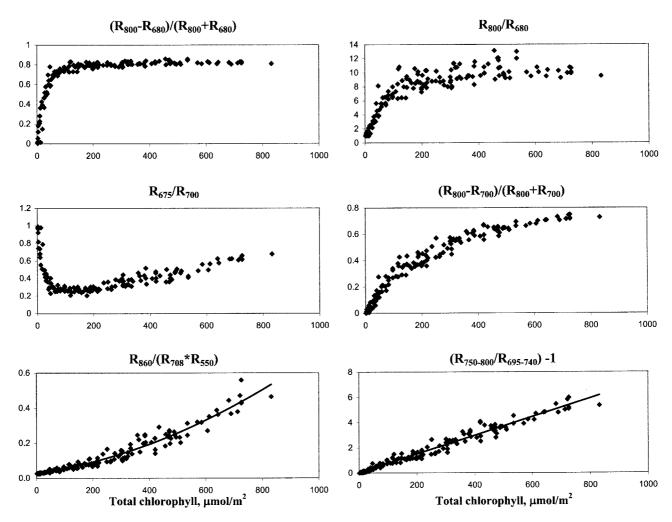


Figure 8. Scatter plots indicating the relationship between various reflectance indices and total ChI content.

(R₆₇₅/R₇₀₀) - Chappelle et al. 1992,

 $(R_{800}-R_{700})/(R_{800}+R_{700})$ – Gitelson and Merzlyak 1994 a and b, $(R_{860}/R_{708} * R_{550})$ – Datt 1998,

 $(R_{750-800})/(R_{695-740})-1$ – this paper.

Indices that used reflectance in the red range around 680 nm were sensitive to only very low ChI and non-sensitive to moderate to high ChI. Thus, the index $(R_{800}-R_{680})/(R_{800}+R_{680})$ became invariant with respect to ChI > $100\,\mu$ mol/m² (Fig. 8 A), and the index (R_{800}/R_{680}) became insensitive to ChI > $200\,\mu$ mol/m² (Fig. 8 B).

The index R_{675}/R_{700} decreased sharply with an increase in total ChI up to $70\,\mu\text{mol/m}^2$, then, it was almost invariant with respect to ChI between 70 and $200\,\mu\text{mol/m}^2$ and had a quite linear relationship with ChI higher than $200\,\mu\text{mol/m}^2$. The indices $[R_{800}-R_{700}]/[R_{800}+R_{700}],$ $[R_{860}/R_{708}*R_{550}]$ and $[(R_{750-800})/(R_{695-740})-1]$ were much better indicators of ChI content. The index $[R_{800}-R_{700}]/[R_{800}+R_{700}]$ was shown to be a good predictor of ChI for a variety of plant species (Gitelson and Merzlyak 1994 a, b, Gamon and Surfus 1999, Richardson et al.

2002, Sims and Gamon 2002), however, the sensitivity of the index to high total ChI (more than $400\,\mu\text{mol/m}^2)$ was four-fold lower than to low-to-moderate ChI. In the range of total ChI variation from 1 to $830\,\mu\text{mol/m}^2$, indices $[R_{800}-R_{700}]/[R_{800}+R_{700}]$ and $[R_{860}/R_{708}*R_{550}]$ provided estimation of total ChI with a RMSE < $80\,\mu\text{mol/m}^2$. Indices with broad spectral bands $[(R_{750-800})/(R_{695-740})\text{-}1]$ and $[(R_{750-800})/(R_{520-585})\text{-}1]$, proposed in this work were the best ChI predictors with a RMSE < $39\,\mu\text{mol/m}^2$ (Table 3).

Three spectral bands, either $550\pm20\,\mathrm{nm}$ or $715\pm20\,\mathrm{nm}$, $450\pm20\,\mathrm{nm}$, and NIR band above $750\,\mathrm{nm}$ were found to be sufficient for non-destructive ChI estimation. For anthocyanin-containing leaves (see Gitelson et al. 2001), spectral bands $715\pm20\,\mathrm{nm}$, $450\pm20\,\mathrm{nm}$ and a NIR band above $750\,\mathrm{nm}$ are recommended. The use of wide band filters in reflectometers for ChI estimation allows for a significant increase in sensitivity and the signal-to-noise ratio and decreases the cost of the reflectometers.

Table 3. Root mean square error of total Chl, Chl *a* and Chl *b* estimation calculated for 142 maple, chestnut, beech and wild vine leaves included in the model development data set. Total Chl content ranged between 1 and 832 μ mol/m², Chl *a* from 1 to 620 μ mol/m² and Chl *b* from 1 to 210 μ mol/m².

RMSE	$(R_{800} - R_{680})/$ $(R_{800} + R_{680})$	R ₇₈₀ /R ₆₈₀	R ₆₇₅ /R ₇₀₀	$(R_{800}-R_{700})/$ $(R_{800}+R_{700})$	R ₈₆₀ / (R ₇₀₈ * R ₅₅₀)	(R ₇₅₀₋₈₀₀ / R ₆₉₅ -740)-1
Chla+b	168	139	63*	80	60	39
Chl a	123	102	49*	58	47	29
Chl b	46	39	20*	24	16	14

^{*} The RMSE of ChI estimation in the range of total ChI > 200 μmol/m², ChI a > 148 μmol/m² and ChI b > 60 μmol/m².

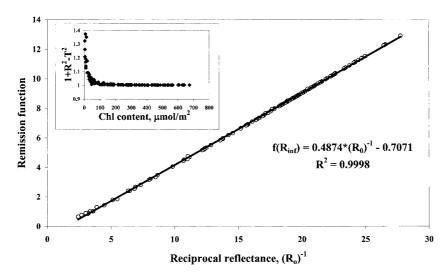


Figure 9. Remission function $f(R_{\infty})$ plotted versus reciprocal reflectance $(R_0)^{-1}$. Linear best-fit function describes this relationship with $r^2 > 0.9998$. In insert: numerator of equation 2 (Appendix 1) is very close to unity in a wide range of total Chl. Both R^2 and T^2 decrease virtually synchronously with an increase in Chl content.

Appendix 1

The relationship between the infinite reflectance of an ideal layer, R_{∞} , in which a further increase in thickness results in no noticeable difference, and inherent optical properties (absorption coefficient, k, and scattering coefficient, s) is as following (e.g., Kortüm 1969):

$$f(R_{\infty}) = (1 - R_{\infty}^{2})/2R_{\infty} = k/s \tag{1}$$

To find the relationship between reflectance of a real leaf and its inherent optical properties, one should find a relationship between $f(R_\infty)$ and the measured leaf reflectance. In our work, a leaf was placed on a black velvet background, and the reflectance was measured using an integrating sphere. In terms of the Kubelka-Munk theory, the measured reflectance is defined as R_o where the index <0> is used to designate the ideal black background with reflectance of the background equal to 0. The relationship between R_0 and $f(R_\infty)$ may be retrieved from the equation (Kortüm 1969, p. 120):

$$f(R_{\infty}) = (1 + R_0^2 - T^2)/2R_0 - 1 \tag{2}$$

where T is leaf transmittance.

The numerator $(1 + R_o^2 - T^2)$ is very close to unity (insert in Fig. 9), and $f(R_\infty)$ is the hyperbolic function of R_o , therefore, $f(R_\infty) \propto (R_o)^{-1}$. For R_o ranging from 0 to 50%, the relationship between $f(R_\infty)$ and $(R_o)^{-1}$ is linear, $f(R_\infty) = 0.4874^*(R)^{-1} - 0.707$ with $r^2 = 0.9998$ (Fig. 9).

Acknowledgements. The authors are very thankful to Professor Harmut K. Lichtenthaler and Dr. Claus Buschmann, Karlsruhe University, for their help in collecting beech and wild vine data and their beneficial discussions. We also acknowledge Yoav Zur for providing us with data on beech 2000. The study was supported partially by a fellowship to Anatoly Gitelson (1996) from the German Academic Exchange Service (DAAD), which is gratefully acknowledged.

References

Adams JE, Arkin GF (1977) A light interception method for measuring row crop ground cover. Soil Sci Soc Amer J 41: 789–792

- Aoki M, Yabuki K, Totsuka T, Nishida M (1986) Remote sensing of chlorophyll content of leaf (I) effective spectral reflection characteristics of leaf for the evaluation of chlorophyll content in leaves of dicotyledons. Environ Control Biol 24: 21–26
- Blackburn GA (1998) Quantifying chlorophylls and carotenoids at leaf and canopy scales: An evaluation of some hyperspectral approaches. Remote Sens Environ 66: 273–285
- Buschmann C, Nagel E (1993) In vivo spectroscopy and internal optics of leaves as basis for remote sensing of vegetation. Int J Remote Sens 14: 711–722
- Carter GA, Knapp AK (2001) Leaf optical properties in higher plants: linking spectral characteristics to stress and chlorophyll concentration. Amer J Bot 84: 677–684
- Chappelle EW, Kim MS, McMurtrey JE, III (1992) Ratio analysis of reflectance spectra (RARS): An algorithm for the remote estimation of the concentrations of chlorophyll *a*, chlorophyll *b*, and carotenoids in soybean leaves. Remote Sens Environ 39: 239–247
- Cui M, Vogelmann TC, Smith WK (1991) Chlorophyll and light gradients in sun and shade leaves of Spinacia oleraceae. Plant Cell Environ 14: 493–500
- Curran PJ, Dungan JL, Gholz HL (1990) Exploring the relationship between reflectance red edge and chlorophyll content in slash pine. Tree Physiol 7: 33–48
- Datt B (1998) Remote sensing of chlorophyll *a*, chlorophyll *b*, chlorophyll *a* + *b*, and total carotenoid content in eucalyptus leaves. Remote Sens Environ 66: 111–121
- Filella I, Serrano I, Serra J, Peñuelas J (1995) Evaluating wheat nitrogen status with canopy relfectance indices and discriminant analysis. Crop Sci 35: 1400–1405
- Fukshansky L (1981) Optical properties of plant tissue. In: Smith H (ed) Plants and the Daylight Spectrum. Springer, Berlin pp 253–303
- Fukshansky LA, Remisowsky AM, McClendon J, Ritterbusch A, Richter T, Mohr H (1993) Absorption spectra of leaves corrected for scattering and distributional error: a radiative transfer and absorption statistics treatment. Photochem Photobiol 57: 538–555
- Gamon JA, Surfus JS (1999) Assessing leaf pigment content and activity with a reflectometer. New Phytol 143: 105–117
- Gates DM, Keegan HJ, Schleter JC, Weidner VR (1965) Spectral properties of plants. Appl Opt 4: 11–20
- Gausman HW (1984) Evaluation of factors causing reflectance differences between sun and shade leaves. Remote Sens Environ 15: 177–181
- Gausman HW, Allen WA (1973) Optical parameters of leaves of 30 plant species. Plant Physiol 52: 57–62
- Gausman HW, Allen WA, Cardenas R (1969) Reflectance of cotton leaves and their structure. Remote Sens Environ 1: 19–22
- Gitelson AA, Merzlyak MN (1994 a) Quantitative estimation of chlorophyll a using reflectance spectra: Experiments with autumn chestnut and maple leaves. J Photochem Photobiol (B) 22: 247–252
- Gitelson AA, Merzlyak MN (1994 b) Spectral reflectance changes associated with autumn senescence of Aesculus hippocastanum and Acer platanoides leaves. Spectral features and relation to chlorophyll estimation. J Plant Physiol 143: 286–292
- Gitelson A, Merzlyak MN (1996) Signature analysis of leaf reflectance spectra: algorithm development for remote sensing of chlorophyll. J Plant Physiol 148: 494–500
- Gitelson A, Merzlyak MN (1997) Remote estimation of chlorophyll content in higher plant leaves. Int J Remote Sens 18: 291–298
- Gitelson AA, Kaufman YJ, Merzlyak MN (1996) Use of a green channel in remote sensing of global vegetation from EOS-MODIS. Remote Sens Environ 58: 289–298

- Gitelson AA, Merzlyak MN, Chivkunova OB (2001) Optical properties and non-destructive estimation of anthocyanin content in plant leaves. Photochem Photobiol 74: 38–45
- Gitelson AA, Zur Y, Chivkunova OB, Merzlyak MN (2002) Assessing carotenoid content in plant leaves with reflectance spectroscopy. Photochem Photobiol 75: 272 281
- Heath OVS (1969) The physiological aspects of photosynthesis. Stanford University Press, Stanford, California, 310 p
- Hendry GAF, Houghton JD, Brown SB (1987) The degradation of chlorophyll biological enigma. New Phytol 107: 255–302
- Kortüm G (1969) Reflectance Spectroscopy. Principles, Methods, Applications. Springer-Verlag, New York Inc
- Kumar R, Silva L (1973) Light ray tracing through a leaf cross section. Appl Optics 12 (12): 2950–2954
- Lichtenthaler HK (1987) Chlorophyll and carotenoids: Pigments of photosynthetic biomembranes. Meth Enzym 148: 331–382
- Lichtenthaler HK, Gitelson AA, Lang M (1996) Non-destructive determination of chlorophyll content of leaves of a green and an aurea mutant of tobacco by reflectance measurements. J Plant Physiol 148: 483–493
- Markwell J, Osterman JC, Mitchell JL (1995) Calibration of the Minolta SPAD-502 leaf chlorophyll meter. Photosynth Res 46: 467–472
- Merzlyak MN, Gitelson AA (1995) Why and what for the leaves are yellow in autumn? On the interpretation of optical spectra of senescing leaves (*Acer platanoides* L.). J Plant Physiol 145: 315–320
- Merzlyak MN, Gitelson AA, Chivkunova OB, Rakitin VY (1999) Nondestructive optical detection of leaf senescence and fruit ripening. Physiol Plant 106: 135–141
- Moran JA, Mitchell AK, Goodmanson G, Stockburger KA (2000) Differentiation among effects of nitrogen fertilization treatments on conifer seedlings by foliar reflectance: a comparison of methods. Tree Physiol 20: 1113–1120
- Moss RA, Loomis WE (1952) Absorption spectra of leaves. I. The visible spectrum. Plant Physiol 27: 370–391
- Peñuelas J, Filella I (1998) Visible and near-infrared reflectance techniques for diagnosing plant physiological status. Trends Plant Sci 3: 151–156
- Richardson AD, Duigan SP, Berlyn GP (2002) An evaluation of noninvasive methods to estimate foliar chlorophyll content. New Phytol 153: 185–194
- Richter T, Fukshansky L (1996) Optics of a bifacial leaf: 2. Light regime as affected by the leaf structure and the light source. Photochem Photobiol 63: 517–527
- Sims DA, Gamon JA (2002) Relationship between leaf pigment content and spectral reflectance across a wide range species, leaf structures and development stages. Remote Sens Environ 81: 337–354
- Thomas JR, Gausman HW (1977) Leaf reflectance vs. leaf chlorophyll and carotenoid concentrations for eight crops. Agron J 69: 799–802
- Vogelmann TC (1993) Plant tissue optics. Annu Rev Plant Physiol Plant Mol Biol 44: 231–251
- Vogelmann TC, Martin G, Chen G, Buttry D (1991) Fibre optic microprobes and measurement of the light microenvironment within plant tissues. Adv Bot Res 18: 231–270
- Wendlandt WWM, Hecht HG (1966) Theory II, in Reflectance Spectroscopy. John Wiley & Sons, New York London Sydney pp 46–90
- Yoder BJ, Pettigrew-Crosby RE (1995) Predicting nitrogen and chlorophyll concentrations from reflectance spectra (400–2500 nm) at leaf and canopy scales. Remote Sens Environ 53: 199–211